

# Intraglomerular Lateral Inhibition Promotes Spike Timing Variability in Principal Neurons of the Olfactory Bulb

Marion Najac,<sup>1,2</sup> Alvaro Sanz Diez,<sup>3</sup>  Arvind Kumar,<sup>4</sup> Nuria Benito,<sup>3</sup> Serge Charpak,<sup>1,2</sup> and  Didier De Saint Jan<sup>1,2,3</sup>

<sup>1</sup>Institut National de la Santé et de la Recherche Médicale Unité 1128, and <sup>2</sup>Université Paris Descartes, 75006 Paris, France, <sup>3</sup>Institut des Neurosciences Cellulaires et Intégratives, Centre National de la Recherche Scientifique, Unité Propre de Recherche 3212, Université de Strasbourg, 67084 Strasbourg, France, and <sup>4</sup>Computational Biology, School of Computer Science and Communication, Royal Institute of Technology, Stockholm, SE-110 40, Sweden

The activity of mitral and tufted cells, the principal neurons of the olfactory bulb, is modulated by several classes of interneurons. Among them, diverse periglomerular (PG) cell types interact with the apical dendrites of mitral and tufted cells inside glomeruli at the first stage of olfactory processing. We used paired recording in olfactory bulb slices and two-photon targeted patch-clamp recording *in vivo* to characterize the properties and connections of a genetically identified population of PG cells expressing enhanced yellow fluorescent protein (EYFP) under the control of the Kv3.1 potassium channel promoter. Kv3.1–EYFP<sup>+</sup> PG cells are axonless and monoglomerular neurons that constitute ~30% of all PG cells and include calbindin-expressing neurons. They respond to an olfactory nerve stimulation with a short barrage of excitatory inputs mediated by mitral, tufted, and external tufted cells, and, in turn, they indiscriminately release GABA onto principal neurons. They are activated by even the weakest olfactory nerve input or by the discharge of a single principal neuron in slices and at each respiration cycle in anesthetized mice. They participate in a fast-onset intraglomerular lateral inhibition between principal neurons from the same glomerulus, a circuit that reduces the firing rate and promotes spike timing variability in mitral cells. Recordings in other PG cell subtypes suggest that this pathway predominates in generating glomerular inhibition. Intraglomerular lateral inhibition may play a key role in olfactory processing by reducing the similarity of principal cells discharge in response to the same incoming input.

**Key words:** glomerulus; inhibition; olfactory bulb; periglomerular cell

## Introduction

The olfactory bulb is the first station in the brain for odor processing (Shepherd, 2004). It receives sensory afferents from olfactory sensory neurons (OSNs) that bind odorants in the nasal epithelium. This information is transmitted to mitral and tufted cells, the principal output neurons of the bulb, within anatomical structures called glomeruli. Olfactory bulb networks modulate how mitral and tufted cells integrate and transform the spatio-temporal map of activated glomeruli into a temporal pattern of action potentials, i.e., the odor code that is ultimately decoded

by downstream cortical neurons. However, the role of bulbar inhibitory circuits in shaping this temporal code is only partly understood.

Several types of interneurons interact with mitral and tufted cells. Granule and parvalbumin-positive (PV<sup>+</sup>) cells make reciprocal synapses with the lateral dendrites of mitral and tufted cells. Mitral and tufted cells activate these interneurons that in turn release GABA onto mitral and tufted cells (Isaacson and Strowbridge, 1998; Schoppa et al., 1998; Christie et al., 2001; Kato et al., 2013; Miyamichi et al., 2013). PV<sup>+</sup> cells mediate a broad and nonselective interglomerular lateral inhibition of widely distributed mitral and tufted cells, whereas granule cells inhibit fewer narrowly confined principal neurons projecting into specific glomeruli (Kato et al., 2013; Miyamichi et al., 2013). Models of olfactory bulb processing stress the importance of interglomerular lateral inhibition for sharpening odor tuning of principal neurons, enhancing contrast between glomerular units and facilitating odor discrimination (Yokoi et al., 1995; Mori et al., 1999; Abraham et al., 2010).

In contrast, little is known about the role of periglomerular (PG) interneurons that surround glomeruli and make synapses onto mitral and tufted cell intraglomerular dendrites (Pinching and Powell, 1971a; Toida et al., 1998; Kosaka and Kosaka, 2007). However, PG cells may provide as much as 50% of their inhibitory inputs (Dong et al., 2007), and recent evidence suggest that a stimulation of OSNs evokes a potent intraglomerular inhibition

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Correspondence should be addressed to either of the following: Didier De Saint Jan, Institut des Neurosciences Cellulaires et Intégratives, Centre National de la Recherche Scientifique, Unité Propre de Recherche 3212, 5, rue Blaise Pascal, 67084 Strasbourg, France. E-mail: desaintjan@inci-cnrs.unistra.fr; or Serge Charpak, Laboratoire de Neurophysiologie et des Nouvelles Microscopies, Institut National de la Santé et de la Recherche Médicale, Unité 1128, 45, rue des Saints Pères, 75006 Paris, France. E-mail: serge.charpak@parisdescartes.fr.

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in mitral cells (Najac et al., 2011; Shao et al., 2012). However, PG cells constitute a diverse population of interneurons with different membrane properties (McQuiston and Katz, 2001; Hayar et al., 2004; Murphy et al., 2005; Shao et al., 2009), morphologies (Pinching and Powell, 1971b; Hayar et al., 2004; Kiyokage et al., 2010; Kosaka and Kosaka, 2010), and immunohistochemical properties (Kosaka and Kosaka, 2007; Panzanelli et al., 2007; Parrish-Aungst et al., 2007; Whitman and Greer, 2007), suggesting that each subtype may form circuits playing different functions.

Thus, to get a better insight into the function of intraglomerular inhibition, we investigated the properties and the synaptic circuits made by a specific subset of PG cells expressing enhanced yellow fluorescent protein (EYFP) under the control of the Kv3.1 potassium channel promoter (Metzger et al., 2002). We determined *in vivo* and *in vitro*, using targeted patch-clamp recording, the circumstances under which labeled PG cells are activated. Our results demonstrate that they generate a prominent intraglomerular lateral inhibition that is activated at each respiration cycle and modulates the frequency and precision of mitral and tufted cell discharges.

## Materials and Methods

**Slice preparation.** Experimental protocols were approved by the National Institute of Health and Medical Research and the National Center of Scientific Research guidelines. Horizontal olfactory bulb slices were prepared from 14- to 30-d-old transgenic mice of either sex expressing the EYFP under the control of the Kv3.1 K<sup>+</sup> channel promoter (Metzger et al., 2002) or the enhanced green fluorescent protein (EGFP) under the control of the calretinin (CR) promoter (Caputi et al., 2009). Note that, in these mice, the fluorescent protein expression may differ from the expression pattern of the Kv3.1 protein (Metzger et al., 2002) or the CR protein (Caputi et al., 2009). Mice were killed by decapitation, and the bulbs were dissected rapidly in ice-cold oxygenated (95% O<sub>2</sub>/5% CO<sub>2</sub>) solution containing the following (in mM): 83 NaCl, 26.2 NaHCO<sub>3</sub>, 1 NaH<sub>2</sub>PO<sub>4</sub>, 2.5 KCl, 3.3 MgSO<sub>4</sub>, 0.5 CaCl<sub>2</sub>, 70 sucrose, and 22 D-glucose, pH 7.3 (osmolality, 300 mOsm/L). Slices (300 μm) were cut using a Leica VT1000S or a Microm HM650V vibratome in the same solution, incubated for 30–40 min at 34°C, and stored at room temperature in a regular artificial CSF (ACSF) until use. ACSF contained the following (in mM): 125 NaCl, 25 NaHCO<sub>3</sub>, 2.5 KCl, 1.25 NaH<sub>2</sub>PO<sub>4</sub>, 1 MgCl<sub>2</sub>, 2 CaCl<sub>2</sub>, and 25 D-glucose (continuously bubbled with 95% O<sub>2</sub>/5% CO<sub>2</sub>).

**Electrophysiological recordings in slices.** Experiments were conducted at 32–34°C on an upright microscope (Olympus BX51WI) with differential interference contrast optics. Whole-cell voltage-clamp recordings of mitral, tufted, and external tufted (ET) cells were made with patch pipettes (~2–4 MΩ) filled with an intracellular solution containing the following (in mM): 120 Cs-MeSO<sub>3</sub>, 20 tetraethylammonium-Cl, 5 4-aminopyridine, 2 MgCl<sub>2</sub>, 0.025 CaCl<sub>2</sub>, 1 EGTA, 4 Na-ATP, 0.5 Na-GTP, and 10 HEPES, pH 7.3 (~280 Osm/L, ~10 mV junction potential). Whole-cell current-clamp recordings were made with an internal solution containing the following (in mM): 135 K-gluconate, 2 MgCl<sub>2</sub>, 0.025 CaCl<sub>2</sub>, 1 EGTA, 4 Na-ATP, 0.5 Na-GTP, and 10 HEPES, pH 7.3 (~280 Osm/L, ~15 mV junction potential). PG cell recordings were all made with pipettes (~4–6 MΩ) filled with the K-gluconate-based internal solution. We discriminated between tufted cells and ET cells as described previously (Najac et al., 2011). Alexa Fluor 594 (6 μM; Invitrogen) was always added to the internal solution to confirm the glomerular projection of the recorded cell. For paired recording of synaptic transmission, the K-gluconate-based internal solution in the presynaptic neuron was supplemented with 10 mM glutamate or 10 mM GABA to avoid transmitter depletion and the resulting rundown of the postsynaptic response. Loose cell-attached recordings (15–100 MΩ seal resistance) were made with pipettes filled with ACSF and used several times on different neurons. Bundles of OSN axons projecting inside a given glomerulus were stimulated using a theta pipette filled with ACSF as described previously (Najac et al., 2011). The electrical stimulus (100 μs) was delivered using an

isolator-11 Stimulus Isolation Unit (Molecular Devices). Recordings were acquired with a multiclamp 700A amplifier (Molecular Devices), low-passed filtered at 2–4 kHz, and digitized at 10 kHz using Clampex 9 software (Molecular Devices). In current-clamp recordings, a constant hyperpolarizing current was injected to maintain the cell at a potential of –60/–70 mV. In voltage-clamp recordings, access resistance ( $R_a$  < 10 MΩ for mitral,  $R_a$  < 20 MΩ for tufted and ET cells,  $R_a$  < 30 MΩ for PG cells) were not compensated. Voltages indicated throughout were corrected for the junction potential estimated with the Clampex software.

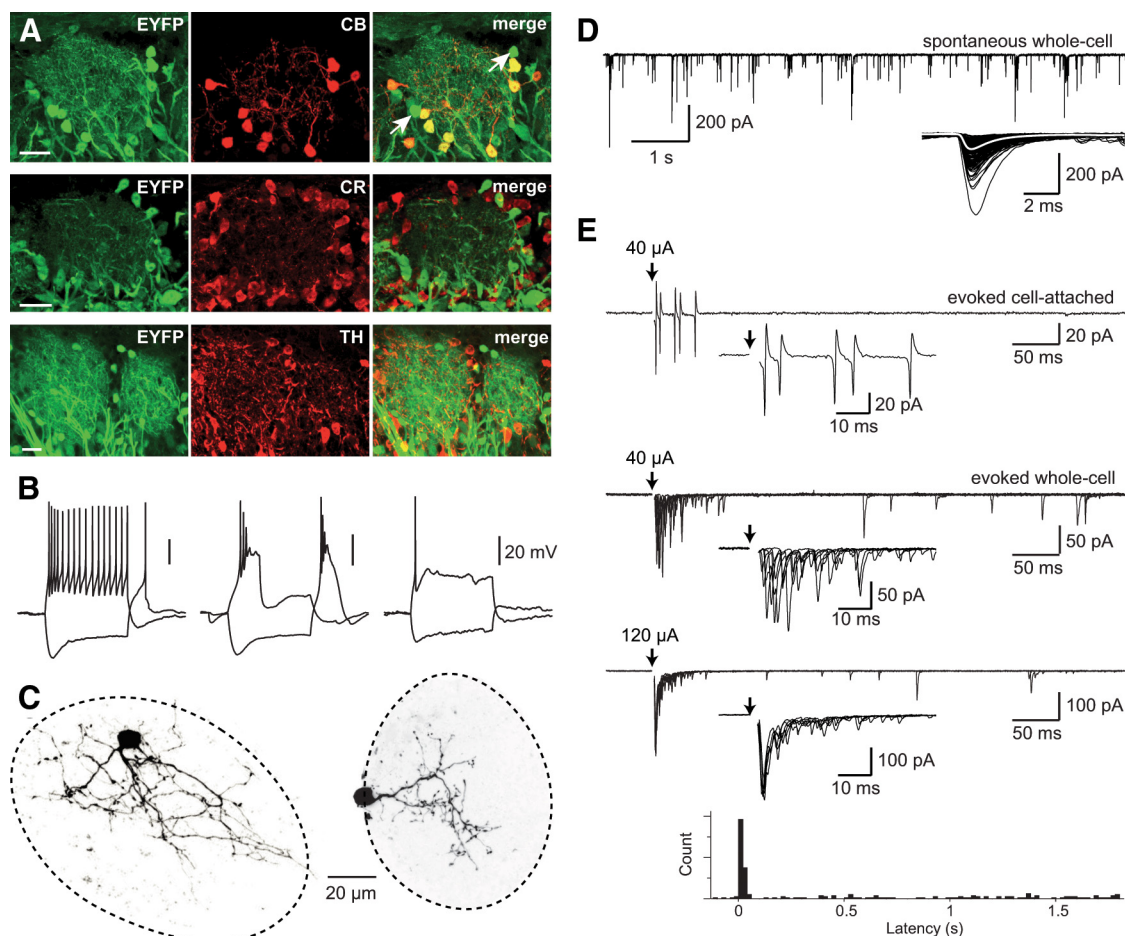
**Drugs.** 6-Nitro-7-sulfamoylbenzo[*f*]quinoxaline-2,3-dione (NBQX), D-2-amino-5-phosphonopentanoic acid (D-AP-5), 2-(3-carboxypropyl)-3-amino-6-(4-methoxyphenyl)pyridinium bromide (GBZ), 7-(hydroxymino)cyclopropa[*b*]chromene-1a-carboxylate ethyl ester (CPCOEt), and 4-ethylphenylamino-1,2-dimethyl-6-methylaminopyrimidinium chloride (ZD7288) were purchased from Tocris Bioscience or Ascent Scientific.

**Intraglomerular application of GBZ.** Local intraglomerular applications of GBZ were made with a patch pipette filled with ACSF supplemented with GBZ. Alexa Fluor 594 (10 μM) was also added to the pipette solution to monitor diffusion with fluorescence. Air pressure at the back of the pipette was applied either transiently (100–200 ms) using an electrical valve or continuously using a syringe.

**PG cell morphology.** To reconstruct the morphology of PG cells, biocytin (10 mM) was added to the intracellular solution, and the patch pipette was slowly withdrawn immediately after breaking the membrane to avoid damaging the cell bodies. Slices were then fixed in 4% paraformaldehyde overnight, washed three times, and incubated in a permeabilizing solution containing Cy5-conjugated streptavidin (1 μg/ml; Jackson ImmunoResearch) for 1 d. After three wash cycles with PBS, sections were mounted. Labeled cells were imaged with a confocal microscope (Zeiss LSM 510).

**Two-photon targeted patch-clamp recordings in vivo.** Six- to 12-week-old Kv3.1-EYFP transgenic mice were anesthetized with an intraperitoneal injection of xylazine (16 mg/kg) and ketamine (100 mg/kg) and placed in a custom stereotaxic apparatus. The posterior cisterna was drained, and a small craniotomy was done on the dorsal side of one olfactory bulb. The dura was removed before stabilizing the bulb with agarose (2% w/v). After surgery, the animal was placed under a custom-built two-photon laser scanning microscope (Chrapak et al., 2001) equipped with a 40× water-immersion objective (numerical aperture, 0.8) and a Mira 900 mode-locked titanium:sapphire laser (Coherent). Anesthesia was switched to isoflurane (1–2%, 3 L/min in moisturized air/O<sub>2</sub>), and its depth was monitored throughout by respiration rates along with lack of response to toe pinch. Body temperature was measured with a rectal probe (FHC) and maintained at 36–37°C with a heating pad (FHC). Respiration was monitored with a pressure pad (Biopac Systems) positioned under the chest. For recordings, we followed the two-photon targeted patching approach (Margrie et al., 2003) using regular patch pipettes (~4–8 MΩ) containing Alexa Fluor 594 (10–20 μM). The dye-filled patch pipette and EYFP<sup>+</sup> neurons were imaged simultaneously with two detection channels using an excitation wavelength of 900 nm. The pipettes were filled with (in mM) 130 K-gluconate, 4 MgCl<sub>2</sub>, 4 Na-ATP, 0.5 Na-GTP, 10 HEPES, and 10 phosphocreatinine, pH 7.3 (~280 Osm/L) for the experiments shown in Figure 6A or with the Cs-MeSO<sub>3</sub>-based internal solution used in slices for the experiments shown in Figure 6B. Data were acquired the same way as in slices using an Axopatch 200B amplifier (Molecular Devices). Access resistances (25–40 MΩ) were not compensated.

**Analysis.** Voltage-clamp data were digitally low-passed filtered offline at 1–2 kHz when necessary and analyzed using Axograph X (Axograph Scientific) and Igor Pro (WaveMetrics). We measured the onset of EPSC and IPSCs at 5% of the first peak of the response. The latency of olfactory nerve (ON)-evoked EPSCs and IPSCs were defined as the time interval between the beginning of the stimulation artifact and the onset of the EPSC/IPSC. The latency of lateral inhibition in Figure 5 was defined as the time interval between the peak of the first action potential and the onset of the average IPSC. Trains of action potentials in the presynaptic cell were first aligned on the first spike of the train before computing the average IPSC. The jitter of EPSCs was defined as the SD of their latencies.



**Figure 1.** Characterization of EYFP<sup>+</sup> PG cells in Kv3.1-EYFP transgenic mice. **A**, Immunofluorescence staining for CB (top), CR (middle), and TH (bottom) in the glomerular layer of Kv3.1-EYFP transgenic mice. Note that all CB<sup>+</sup> PG cells express EYFP, but some EYFP<sup>+</sup> PG cells do not express CB (arrows). Scale bars, 20  $\mu$ m. **B**, Intrinsic membrane properties were diverse in EYFP<sup>+</sup> PG cells. Depolarizing and hyperpolarizing steps (500 ms) were injected in EYFP<sup>+</sup> PG cells recorded in current-clamp at a membrane potential of approximately  $-60$  mV. The three examples show individual traces from three cells responding differently to a step depolarization. **C**, Two biocytin-filled EYFP<sup>+</sup> PG cells. Dashed lines delimit the glomerulus into which each PG cell projected its dendrites. **D**, Spontaneous EPSCs in an EYFP<sup>+</sup> PG cell recorded at  $V_h = -75$  mV. Onset, Individual spontaneous EPSCs (black traces) and their average (white trace) are superimposed. **E**, Responses evoked by stimulation of OSNs (arrow) recorded in an EYFP<sup>+</sup> PG cell in the cell-attached (top) or whole-cell voltage-clamp mode at two intensities of stimulation indicated on the left (bottom;  $V_h = -75$  mV). Individual traces are superimposed. Insets are zooms on the initial part of the response. Note that the latency and jitter of the responses decreased with the intensity of stimulation. The histogram plots the timing of individual EPSCs in responses evoked by the highest intensity of stimulation (bin width, 20 ms; stimulation at  $t = 0$ ).

Peristimulus time histograms (PSTHs) represent the average number of EPSCs (Fig. 1) or spikes (see Fig. 8) per bin during a sweep or during a respiration cycle (see Fig. 6). Spike timings were detected automatically with NeuroMatic (<http://www.neuromatic.thinkrandom.com/>) by a  $y$ -threshold level detector for values on a positive slope deflection. Reliability of mitral cell spiking responses (see Fig. 8) was analyzed using MATLAB (MathWorks). For the ensemble analysis of mitral cell responses to the stimulation of OSNs (see Fig. 8G), we constructed a nine-neuron ensemble by pooling the nine recorded mitral cells, each of them having at least 14 trials for each condition, with and without GBZ. By random non-redundant combination of nine trials of  $9 \times 14$ , we created 14 artificial trials of the activity of an ensemble of nine mitral cells.

Unless indicated otherwise, we used the unpaired  $t$  test to compare two sets of data acquired from different populations. Results are expressed as mean  $\pm$  SD.

**Immunohistochemistry.** Adult mice (1–3 months old) were deeply anesthetized with an intraperitoneal injection of xylazine (16 mg/kg) and ketamine (100 mg/kg) and perfused with 4% paraformaldehyde in which the brains were kept overnight. Slices were cut with either a vibratome (50  $\mu$ m) or a cryostat (20  $\mu$ m). Before slicing with the cryostat, brains were cryoprotected and frozen. Blocking steps were performed by PBS solution containing 2% BSA and 0.1% Triton X-100. Sections were incubated 48 h at 4°C with primary antibodies: mouse monoclonal anti-

calbindin D-28k (CB; 1:1000; Swant), rabbit anti-CR 7699/4 (1:1000; Swant), or rabbit polyclonal anti-tyrosine hydroxylase (TH; 1:500; Millipore). After washing, they were incubated 2 h at room temperature with secondary antibodies: Alexa Fluor 594-conjugated anti-mouse or anti-rabbit (1:300–1:500; Invitrogen). After washing, sections were mounted in Vectashield (Vector Laboratories). Confocal images were taken using a Zeiss LSM 510 confocal microscope, and cells were counted manually.

## Results

### Morphology, immunohistochemical identity, and membrane properties of a genetically identified PG cell subtype

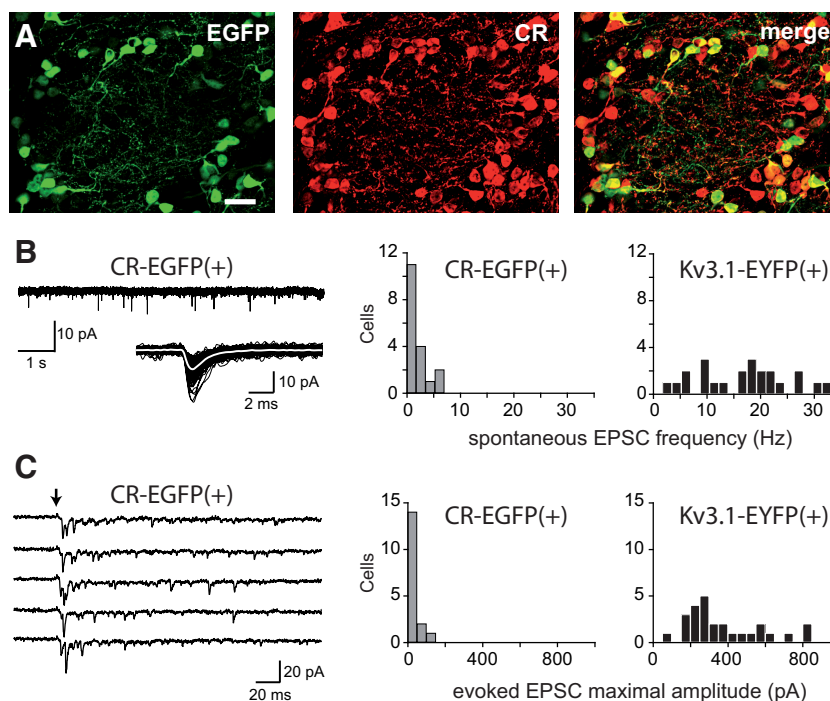
In Kv3.1-EYFP transgenic mice, mitral and tufted cells, as well as a population of juxtaglomerular cells, express EYFP (Metzger et al., 2002). Using slices from juvenile olfactory bulb (15–30 postnatal days), we identified the EYFP<sup>+</sup> juxtaglomerular cells as PG cells (hereafter called EYFP<sup>+</sup> PG cells) on the basis of the small size of their round soma, their localization around glomeruli (Fig. 1A), and their large input resistance ( $950 \pm 375$  M $\Omega$ ,  $n = 20$ ). EYFP<sup>+</sup> PG cells filled with biocytin apparently had no axon ( $n = 11$ ), and 10 of 11 projected their dendrites in a single glomerulus (Fig. 1C). PG cells have been classified anatomically into



two broad classes (Kosaka and Kosaka, 2007): so-called type 1 PG cells are synaptically contacted by OSN axon terminals, whereas type 2 PG cells are not and receive excitatory inputs from the dendrites of mitral, tufted, or ET cells. Prototypical type 2 PG cells express the calcium binding proteins CB or CR (Kosaka and Kosaka, 2007). In Kv3.1-EYFP transgenic mice, virtually all PG cells immunoreactive for CB were EYFP<sup>+</sup> (388 of 392 cells,  $n = 5$  mice), but only half of EYFP<sup>+</sup> PG cells expressed CB (388 of 695 cells; Fig. 1A). EYFP<sup>+</sup> PG cells did not overlap with cells that express CR ( $n = 4$  mice; Fig. 1A) nor with dopaminergic-GABAergic cells expressing TH ( $n = 4$  mice; Fig. 1A). Thus, the population of EYFP<sup>+</sup> PG cells encompasses cells with different immunohistochemical properties. Because CB<sup>+</sup> neurons constitute 14% of all PG cells (Panzanelli et al., 2007), we estimate that ~30% of all PG cells are EYFP<sup>+</sup> in Kv3.1-EYFP mice. Intrinsic membrane properties and firing patterns of EYFP<sup>+</sup> PG cells were also diverse (Fig. 1B). The majority of the cells responded to depolarizing current steps with large calcium spikes ( $n = 16$ ) and the others with sustained firing of sodium action potentials ( $n = 8$ ) or with few sodium spikes preceding a plateau potential ( $n = 4$ ). In most ( $n = 27$  of 28) EYFP<sup>+</sup> PG cells, hyperpolarizing current steps caused a voltage sag. This rebound was blocked by ZD7288 (50  $\mu$ M), an antagonist of  $I_h$ -mediating hyperpolarization-activated cyclic nucleotide gated channels ( $n = 7$  cells; data not shown).

### Synaptic properties of EYFP<sup>+</sup> PG cells

To characterize the synaptic properties of EYFP<sup>+</sup> PG cells, we first examined their spontaneous excitatory synaptic inputs and their responses to a brief (100  $\mu$ s) electrical stimulation of an OSN axon bundle converging into their glomerulus. Whole-cell voltage-clamp recordings of EYFP<sup>+</sup> PG cells ( $V_h = -75$  mV) revealed spontaneous EPSCs at a rate of  $17.5 \pm 8.9$  per second ( $n = 24$ ; Fig. 1D). The average EPSC had an amplitude of  $50 \pm 23$  pA and a fast time course (20–80% rise time,  $0.28 \pm 0.05$  ms; decay time constant,  $1.48 \pm 0.2$  ms). In the cell-attached configuration preceding whole-cell recordings, ON stimulation evoked a burst of few action potentials usually restricted to the first 50 ms after the stimulus ( $3.7 \pm 1.4$  spikes,  $94 \pm 6\%$  within this temporal window,  $n = 14$ ; Fig. 1E). The first spike of the burst occurred with an average latency of  $6.8 \pm 3.1$  ms (range of 3–13.7 ms,  $n = 14$ ) that decreased with increasing stimulation intensity (data not shown). In the whole-cell configuration, ON stimulation evoked a brief barrage of fast summing EPSCs that summed up to  $424 \pm 302$  pA ( $n = 31$ ) at strong intensities of stimulation ( $153 \pm 107$   $\mu$ A; Fig. 1E). To quantify the duration of these responses, we constructed PSTHs of EPSCs and fitted the decay of the histogram with an exponential. As in the example shown in Figure 1E, the majority of the cells ( $n = 26$  of 31) had a response that quickly returned to baseline with a time constant  $<100$  ms (mean of  $64 \pm 66$  ms; see Fig. 3A). At  $V_h = -75$  mV, the AMPA receptor antag-



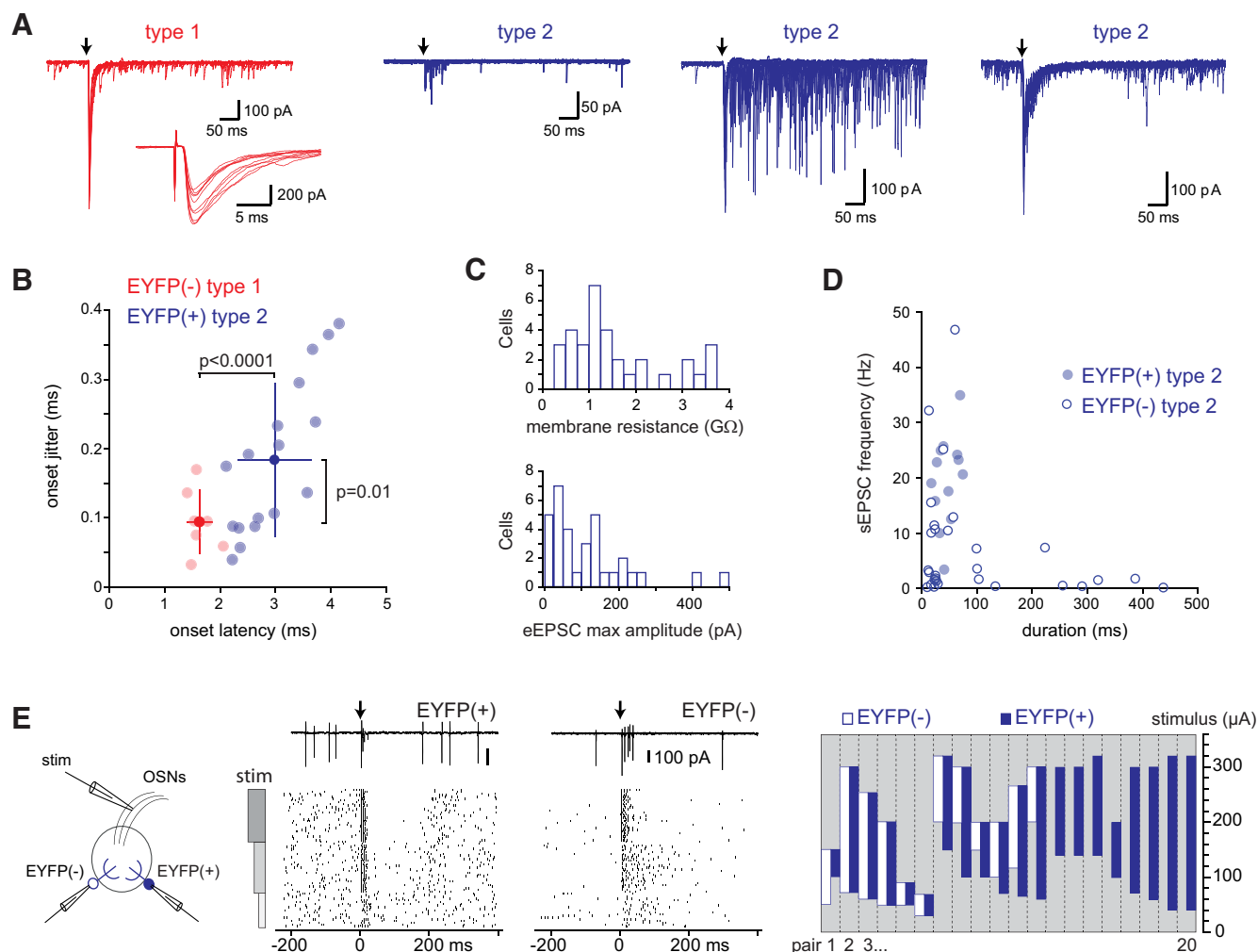
**Figure 2.** A large fraction of CR<sup>+</sup> PG cells are weakly connected to the glomerular network. **A**, Immunofluorescence staining for CR (red, middle) revealed that only a fraction of CR<sup>+</sup> PG cells express EGFP (green, left) in CR-EGFP transgenic mice. Scale bar, 20  $\mu$ m. **B**, Spontaneous EPSCs recorded in the whole-cell voltage-clamp mode ( $V_h = -75$  mV) in a CR-EGFP<sup>+</sup> PG cell. Onset, Individual events (black) superimposed with the average EPSC (white). Distribution histograms compare the frequency of spontaneous EPSCs in CR-EGFP<sup>+</sup> PG cells (left) and Kv3.1-EYFP<sup>+</sup> PG cells (right). **C**, Five consecutive ON-evoked whole-cell responses ( $V_h = -75$  mV) in a CR-EGFP<sup>+</sup> PG cell. This stimulus (200  $\mu$ A, arrow) produced the largest responses. Distribution histograms compare the maximal amplitudes of ON-evoked responses in CR-EGFP<sup>+</sup> PG cells (left) and Kv3.1-EYFP<sup>+</sup> PG cells (right).

onist NBQX (10  $\mu$ M) abolished evoked EPSCs ( $n = 3$ ; data not shown), whereas at  $V_h = 30$  mV, addition of the NMDA receptor antagonist d-AP-5 (50  $\mu$ M) was required to block the EPSC ( $n = 4$ ; data not shown). As reported previously (Shao et al., 2009), the latency of the first EPSC and the jitter of the latency depended on the intensity of stimulation (Fig. 1E). A strong stimulus ( $110 \pm 101$   $\mu$ A) evoked an EPSC with an average latency of  $3 \pm 0.7$  ms and a jitter of  $0.18 \pm 0.11$  ms ( $n = 17$ ). With a weak stimulus ( $36 \pm 14$   $\mu$ A), the average latency was  $4.4 \pm 1.1$  ms and the jitter was  $0.6 \pm 0.5$  ms ( $n = 17$ ). However, the latencies and jitters of responses evoked by stronger intensities were significantly larger than those of monosynaptically connected ET cells (latency,  $2.0 \pm 0.4$  ms,  $p < 0.0001$ ; jitter,  $0.07 \pm 0.06$  ms,  $p = 0.0007$ ;  $n = 12$  ET cells; data not shown) and those of PG cells type 1 (see Fig. 3B) and were consistent with a polysynaptic activation.

### PG cell functional diversity

To evaluate the functional relevance of EYFP<sup>+</sup> PG cells, we compared their ON-evoked activation patterns with those of other PG cell subtypes. PG cells expressing CR are three times more numerous than CB-expressing PG cells and constitute the most abundant immunohistochemical subtype (Panzanelli et al., 2007; Parrish-Aungst et al., 2007; Whitman and Greer, 2007). We used transgenic mice expressing EGFP under the control of the CR promoter to identify these PG cells (Caputi et al., 2009). In CR-EGFP mice, only 56% of the cells immunoreactive for CR were also EGFP<sup>+</sup> (1173 of 2098 cells), but almost all EGFP<sup>+</sup> PG cells were immunoreactive for CR (1173 of 1185 cells,  $n = 6$  mice; Fig. 2A). In the whole-cell configuration, CR-EGFP<sup>+</sup> PG cells had a high input resistance ( $2 \pm 1.3$  G $\Omega$ ,  $n = 30$ ) and received significantly less spontaneous EPSCs ( $1.8 \pm 1.9$  per second,  $n = 18$ )





**Figure 3.** PG cell functional diversity. **A**, ON stimulation (arrow) evoked different types of responses in EYFP<sup>−</sup> PG cells. Type 1 PG cells (red traces) responded with a fast-onset monosynaptic EPSC (the inset is a zoom on the initial part of the response), whereas type 2 PG cells responded with plurisynaptic responses of variable amplitudes and durations (3 examples shown with blue traces).  $V_h = -75$  mV. In each case, several traces are superimposed. **B**, The onset latency versus onset jitter of individual ON-evoked responses is plotted for EYFP<sup>+</sup> PG cells (blue circles) and EYFP<sup>−</sup> type 1 PG cells (red circles). Dark symbols and bars show the mean  $\pm$  SD. Strong intensities of stimulation were used here ( $110 \pm 101$   $\mu$ A for EYFP<sup>+</sup> PGs and  $239 \pm 122$   $\mu$ A for EYFP<sup>−</sup> PGs). **C**, Distribution histograms of the membrane resistance (top) and maximal amplitude of ON-evoked responses (eEPSC; bottom) in EYFP<sup>−</sup> type 2 PG cells ( $n = 33$ ). **D**, The duration of the ON-evoked response is plotted against the frequency of spontaneous EPSCs for each individual EYFP<sup>−</sup> type 2 PG cells (white circles). Values for EYFP<sup>+</sup> PG cells (blue circles) are also plotted for comparison. Durations were estimated from the fit of the latency histogram of each cell. **E**, Comparison of ON-evoked firing in EYFP<sup>+</sup> and EYFP<sup>−</sup> PG cells. Action potentials were monitored using paired loose cell-attached recordings (scheme). Middle, Example of a pair that responded to ON stimulation with a similar threshold but different time course. One representative trace (top) and raster plots of 80 consecutive episodes (bottom) are shown. Each point indicates a spike. OSNs were stimulated (arrow) with increasing intensities. Right, Summary graph of the experiment. Each column shows a pair. White and blue vertical bars show the range of stimulus intensities that evoked the firing of the EYFP<sup>−</sup> and EYFP<sup>+</sup> PG cells, respectively. Columns with no white bars (pairs 13–20) indicate pairs in which the EYFP<sup>−</sup> PG cells did not respond to the stimuli tested.

than EYFP<sup>+</sup> PG cells in the Kv3.1–EYFP mouse ( $p < 0.0001$ ; Fig. 2B). Their average spontaneous EPSC was fast (20–80% rise time,  $0.29 \pm 0.08$  ms; decay,  $1.5 \pm 0.5$  ms) but of smaller amplitude than in EYFP<sup>+</sup> PG cells ( $13.9 \pm 6.3$  pA,  $p < 0.0001$ ). ON stimulation evoked a barrage of EPSCs that summed up to  $33 \pm 34$  pA ( $n = 17$ ) at maximal intensities of stimulation (320  $\mu$ A), a much smaller amplitude than responses evoked in EYFP<sup>+</sup> PG cells ( $p < 0.0001$ ; Fig. 2C). Small inputs could be sufficient to trigger the firing of a neuron with a high membrane resistance. However, most CR–EGFP<sup>+</sup> PG cells did not fire any action potentials spontaneously or in response to ON stimulation in the cell-attached (four of six silent cells) or loose cell-attached (six of eight silent cells) configuration. In the remaining four CR–EGFP<sup>+</sup> cells, the stimulation evoked only one spike (data not shown). Similarly, in paired recording in the loose cell-attached mode of a CR–EGFP<sup>+</sup> and a CR–EGFP-negative (CR–EGFP<sup>−</sup>)

PG cell surrounding the same glomerulus, ON stimulation (50–200  $\mu$ A) that induced the firing of the nonlabeled PG cell failed to activate the CR–EGFP<sup>+</sup> PG cell (in 9 of 11 pairs) or at most elicited a single spike ( $n = 2$  of 11 pairs; data not shown). Thus, a large and previously undescribed fraction of CR<sup>+</sup> PG cells receives little excitatory inputs from the glomerular network and is not, or only weakly, activated by ON inputs that potentially activate other PG cell subtypes.

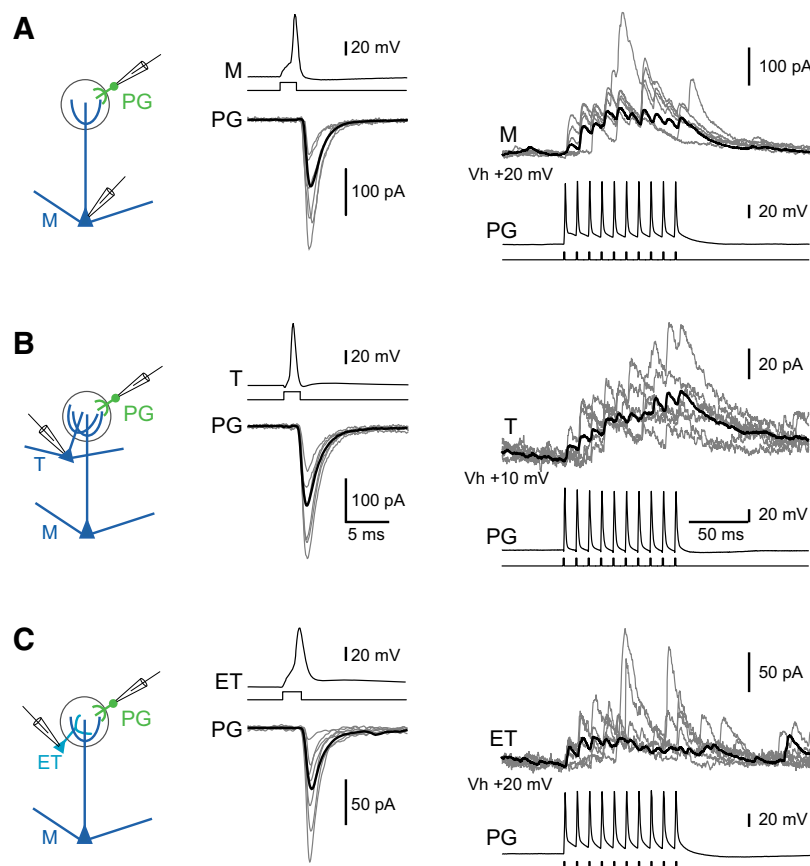
To further explore PG cell diversity, we next examined ON-evoked whole-cell voltage-clamp excitatory responses of randomly chosen nonfluorescent PG cells in Kv3.1–EYFP mice. We identified 17.5% of type 1 PG cells ( $n = 7$  of 40; Fig. 3A) that responded with a fast-onset monosynaptic EPSC whose latency ( $1.64 \pm 0.22$  ms) and jitter ( $0.09 \pm 0.05$  ms) were significantly smaller than those of EYFP<sup>+</sup> PG cells (Fig. 3B). The other EYFP<sup>−</sup> PG cells ( $n = 33$ ) responded with a plurisynaptic barrage of EP-

SCs of variable duration and amplitude. As expected, a large fraction of these type 2 PG cells ( $n = 13$ ) had properties consistent with those of CR-EGFP<sup>+</sup> PG cells: a high membrane resistance ( $>1.5\text{ G}\Omega$ ), small ON-evoked responses ( $<100\text{ pA}$ ) at maximal stimulation intensities, and a low frequency of spontaneous EPSCs ( $<2\text{ Hz}$ ; Fig. 3*A,C*). The remaining EYFP<sup>−</sup> type 2 PG cells ( $n = 20$ ) responded either with a long-lasting barrage of EPSCs that could last several hundred milliseconds (11 cells with ON-evoked responses  $>100\text{ ms}$ ) or with a short plurisynaptic response indistinguishable from those of EYFP<sup>+</sup> PG cells (Fig. 3*A*). Cells with long-lasting responses usually received a lower frequency of spontaneous EPSCs than cells with short responses (Fig. 3*D*). These different types of responses were neither glomerulus specific nor dependent on the stimulation strength because they were also observed in paired recordings between cells projecting into the same glomerulus ( $n = 9$ ; data not shown). Therefore, they rather reflect specific synaptic connectivity and possibly distinguish functionally distinct subtypes.

In a third set of experiments in Kv3.1–EYFP mice, we made paired recordings in the loose cell-attached mode of an EYFP<sup>+</sup> and an EYFP<sup>−</sup> PG cell surrounding the same glomerulus and stimulated OSNs with increasing intensities until threshold ( $n = 20$  pairs; Fig. 3*E*). Seventy percent of EYFP<sup>−</sup> PG cells were not activated by weak stimuli that drive EYFP<sup>+</sup> PG cells. Among these, 40% could not be activated at any intensities of stimulation, and 30% had a higher firing threshold than EYFP<sup>+</sup> PG cells. Most nonresponding cells were totally silent (five of eight), such as CR-EGFP<sup>+</sup> PG cells. The remaining 25% of EYFP<sup>−</sup> PG cells (5 of 20) had a threshold that was approximately similar to EYFP<sup>+</sup> PG cells. Last, only one EYFP<sup>−</sup> PG cell (5%) had a lower threshold than its paired EYFP<sup>+</sup> PG cell. Figure 3*E* (middle) illustrates one pair of PG cells with a similar threshold but different firing patterns: the EYFP<sup>+</sup> PG cell responded as usual with a brief burst of action potentials, followed by a pause during which the EYFP<sup>−</sup> PG cell continued to fire. Thus, although EYFP<sup>+</sup> PG cells are not the only PG cell subtype activated by weak ON inputs, they form a large group of neurons with similar synaptic connections that likely plays a dominant role in generating intraglomerular inhibition in response to weak OSN inputs.

### Synaptic circuits formed by EYFP<sup>+</sup> PG cells

We used paired recordings to identify the presynaptic partners of EYFP<sup>+</sup> PG cells. Three glutamatergic cell types with glomerular projections—mitral, tufted, and ET cells—might provide excitatory inputs onto EYFP<sup>+</sup> PG cells. Contrasting with the idea that type 2 PG cells receive their excitatory input principally from ET cells (Shao et al., 2009), we found that action potentials evoked in any of these glutamatergic neurons produced fast EPSCs in EYFP<sup>+</sup> PG cells (Fig. 4). EPSC amplitudes varied between pairs

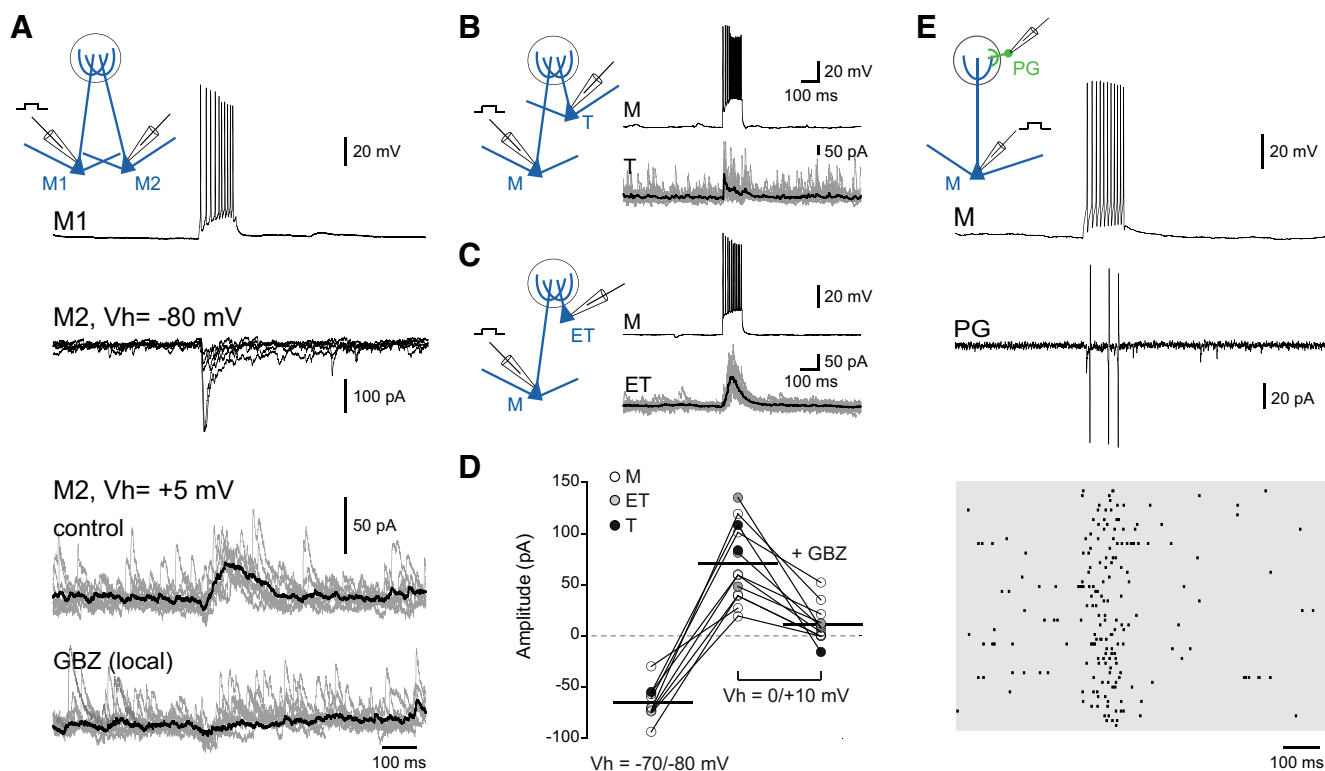


**Figure 4.** Synaptic connections of EYFP<sup>+</sup> PG cells. **A**, Paired recordings between mitral (M) and EYFP<sup>+</sup> PG cells. Left, Single action potentials evoked in the mitral cell produced fast EPSCs in the PG cell recorded at  $V_h = -70\text{ mV}$ . Six representative EPSCs are superimposed in gray, and the average EPSC across all trials is shown in black. Right, In another pair, a train of 10 action potentials evoked in the PG cell produced a barrage of IPSCs in the mitral cell recorded at  $V_h = 20\text{ mV}$ . Six sweeps are superimposed, and the average from all trials appears in black. **B**, Same for connected pairs of tufted (T) and EYFP<sup>+</sup> PG cells. **C**, Same for connected pairs of ET and EYFP<sup>+</sup> PG cells. IPSCs in **A** and **B** were recorded in the presence of NBQX ( $10\text{ }\mu\text{M}$ ) and D-AP-5 ( $50\text{ }\mu\text{M}$ ); **C** in ACSF.

and from trials to trials within pairs. On average, the EPSC amplitude was  $140 \pm 222\text{ pA}$  in ET–PG pairs ( $n = 15$ ),  $124 \pm 57\text{ pA}$  in mitral–PG pairs ( $n = 6$ ), and  $160 \pm 122\text{ pA}$  in tufted–PG pairs ( $n = 5$ ) at  $V_h = -75\text{ mV}$ . In different pairs with mitral/tufted cells filled with a Cs<sup>2+</sup>-based internal solution and voltage clamped around the reversal potential for excitation ( $V_h = 0/20\text{ mV}$ ), 100 Hz trains of action potentials in the EYFP<sup>+</sup> PG cell produced a barrage of summing IPSCs in mitral ( $n = 2$  pairs; Fig. 4*A*), tufted ( $n = 4$ ; Fig. 4*B*), and ET ( $n = 3$ ; Fig. 4*C*) cells. These IPSCs persisted in the presence of NBQX and D-AP-5 ( $n = 7$ ) and were blocked by bath application of GBZ ( $2\text{ }\mu\text{M}$ ,  $n = 5$ ; data not shown). In most pairs (eight of nine), failures of GABAergic transmission were frequent after the first action potential ( $64 \pm 18\%$  of failure). Thus, EYFP<sup>+</sup> PG cells are activated by mitral, tufted, and ET cells and in turn release GABA with a low probability onto mitral, tufted, and ET cells.

### Intraglomerular lateral inhibition between principal cells projecting into the same glomerulus

We predicted that synaptic circuits made by EYFP<sup>+</sup> PG cells may generate lateral inhibition between principal neurons projecting into the same glomerulus (Urban and Sakmann, 2002). Thus, we tested whether principal cells with apical dendrites in the same glomerulus inhibit each other. In pairs of mitral cells, trains of action potentials were evoked in the first cell (Fig. 5, M1), whereas the postsynaptic response of the second cell (M2) was recorded in



**Figure 5.** EYFP<sup>+</sup> PG cells mediate lateral inhibition between mitral, tufted, and ET cells projecting into the same glomerulus. **A**, Paired recording between two mitral cells (M1 and M2) projecting into the same glomerulus. A current step (700 pA, 100 ms) was used to evoke a train of action potentials in M1 (top trace) while simultaneously monitoring voltage-clamped responses in M2 (scheme). Inward EPSCs were recorded at  $V_h = -80$  mV (top traces, 5 traces superimposed). Outward IPSCs were recorded at  $V_h = 5$  mV (bottom gray traces) in control conditions and after a transient puff of GBZ (50  $\mu$ M, 100 ms) within the glomerulus. This protocol was repeated eight times. Traces before (control) and after (GBZ) the puff are superimposed. Average traces are shown in black. **B**, Same experiment in a pair of mitral (M) cell–tufted (T) cell. **C**, Same experiment in a pair of mitral (M) cell–ET cell. Experiments in **A–C** in the presence of D-AP-5 (50  $\mu$ M) and CPCCOEt (100  $\mu$ M). **D**, Summary plot showing the amplitudes of the averaged postsynaptic current obtained at negative potentials ( $V_h = -70/-80$  mV) and at a potential near 0 mV, under control conditions and in the presence of GBZ. Each plot is a mitral (white), tufted (black), or ET (gray) cell. Horizontal lines show the means. **E**, Paired recording between a mitral cell (M) recorded in the whole-cell mode and an EYFP<sup>+</sup> PG cell recorded in the loose cell-attached mode (PG). Both neurons projected into the same glomerulus. A train of action potentials induced by a current step (400 pA, 100 ms) in the mitral cell (top trace) produced the firing of the PG cell (middle trace). Bottom, Raster plots of spontaneous and evoked action potentials in the PG cell.

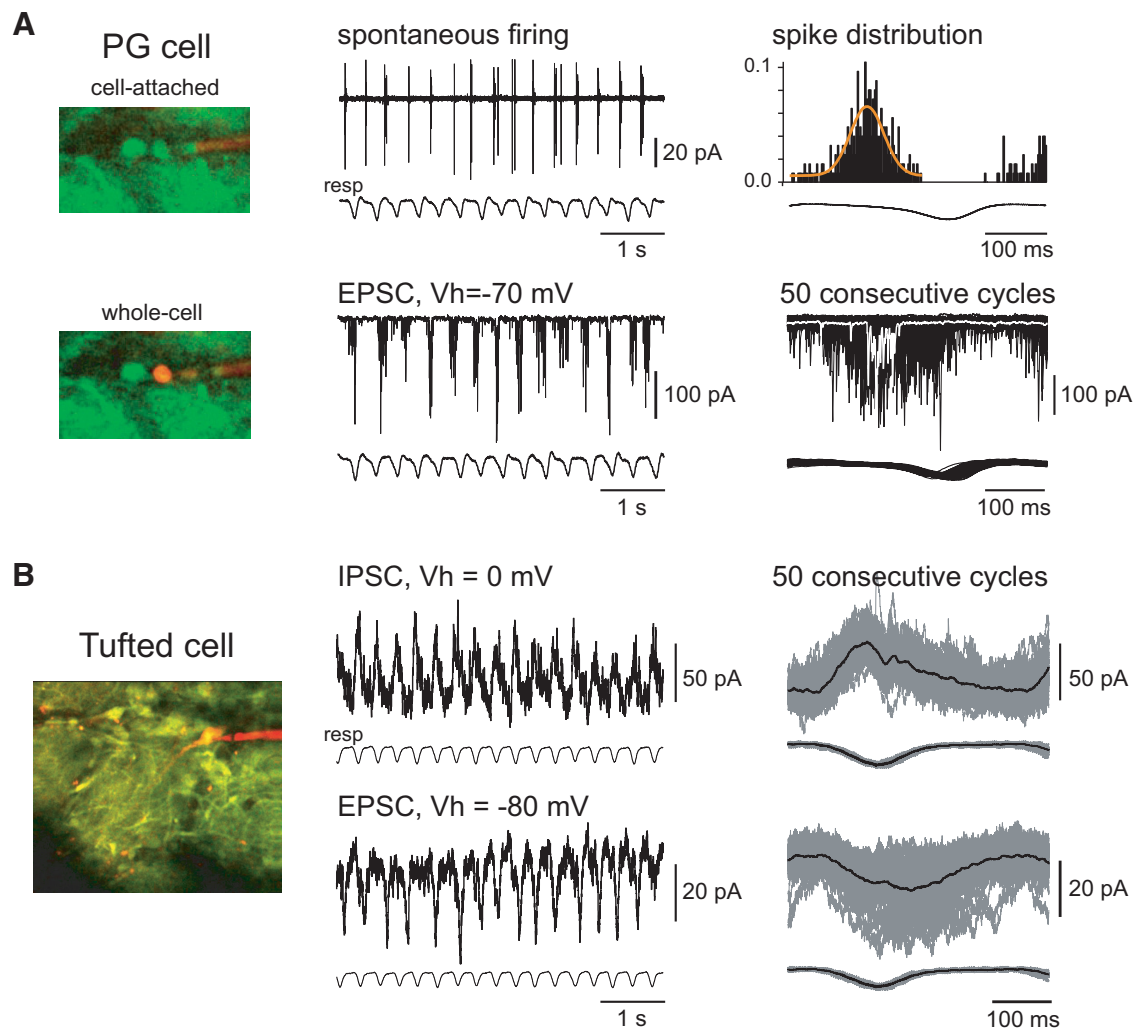
voltage-clamp. These experiments were done in the presence of mGlu1 and NMDA receptor antagonists (CPCCOEt and D-AP-5, respectively) (1) to reduce the excitability of the network and spontaneous synaptic activity and (2) to limit the activation of granule cells. Fast synaptic inward currents were evoked when the postsynaptic cell was recorded at  $V_h = -70/-80$  mV (Fig. 5A,D), consistent with the existence of excitatory synaptic interactions and gap junction coupling between principal neurons affiliated with the same glomerulus (Schoppa and Westbrook, 2002; Urban and Sakmann, 2002). When the postsynaptic mitral cell was voltage clamped around the reversal potential for excitation ( $V_h = 0/10$  mV), a burst of synaptic outward currents was evoked. The average outward current started  $20 \pm 17$  ms (range of 3–50 ms) after the first spike of the presynaptic mitral cell, had an amplitude of  $58 \pm 35$  pA ( $n = 8$ ; Fig. 5D), and was blocked by a transient puff ( $n = 3$ , 100 ms, 50  $\mu$ M) or by a local perfusion of GBZ ( $n = 2$ , 25  $\mu$ M) inside the glomerulus (Fig. 5A,D). Importantly, spontaneous IPSCs persisted during intraglomerular application of GBZ, indicating that the spread of GBZ outside the glomerulus was spatially limited (see Fig. 7). Firing of a single mitral cell also evoked a barrage of polysynaptic IPSCs in tufted cells ( $n = 2$  pairs; amplitude, 83 and 108 pA; onset, 4.3 and 2.9 ms; Fig. 5B) and ET cells (amplitude,  $68 \pm 49$  pA; onset,  $20.7 \pm 35$  ms; range of 2.8–74 ms;  $n = 4$  pairs; Fig. 5C). The polysynaptic inhibition of ET cells is likely generated by intraglomerular circuits because ET cells, that lack lateral dendrites in the external

plexiform layer (EPL), presumably do not interact with granule cells or PV<sup>+</sup> interneurons. Thus, our data suggest that glomerular circuits mediate lateral inhibition between principal neurons projecting into the same glomerulus. Do EYFP<sup>+</sup> PG cells participate in this form of lateral inhibition? If so, they should be activated by the firing of a single mitral cell. We tested this hypothesis in pairs of mitral and EYFP<sup>+</sup> PG cells projecting into the same glomerulus. Trains of action potentials in a mitral cell evoked  $1.8 \pm 0.9$  action potentials in an EYFP<sup>+</sup> PG cell recorded simultaneously in the loose cell-attached configuration in control conditions ( $n = 4$  pairs; Fig. 5E), as well as in the presence of D-AP-5 ( $n = 3$ ; data not shown). Thus, EYFP<sup>+</sup> PG cells participate to the circuit mediating intraglomerular lateral inhibition.

#### Intraglomerular lateral inhibition is highly active *in vivo*

The time course of olfactory processing is constrained by the 2–10 Hz respiratory rhythm. We used two-photon guided targeted patch-clamp recording in freely breathing anesthetized mice to examine the activity of EYFP<sup>+</sup> PG cells *in vivo*. Each recorded EYFP<sup>+</sup> PG cells exhibited rhythmic bursts of fast EPSCs ( $n = 9$ ), consistent with mitral or tufted cell excitatory inputs. These bursts were correlated with the respiration cycle in eight of nine cells as in the example shown in Figure 6A. Moreover, EYFP<sup>+</sup> PG recorded in cell-attached mode before breaking the membrane also fired bursts of action potentials at a specific phase of the respiration cycle ( $n = 5$  of 5 cells; Fig. 6A). These results





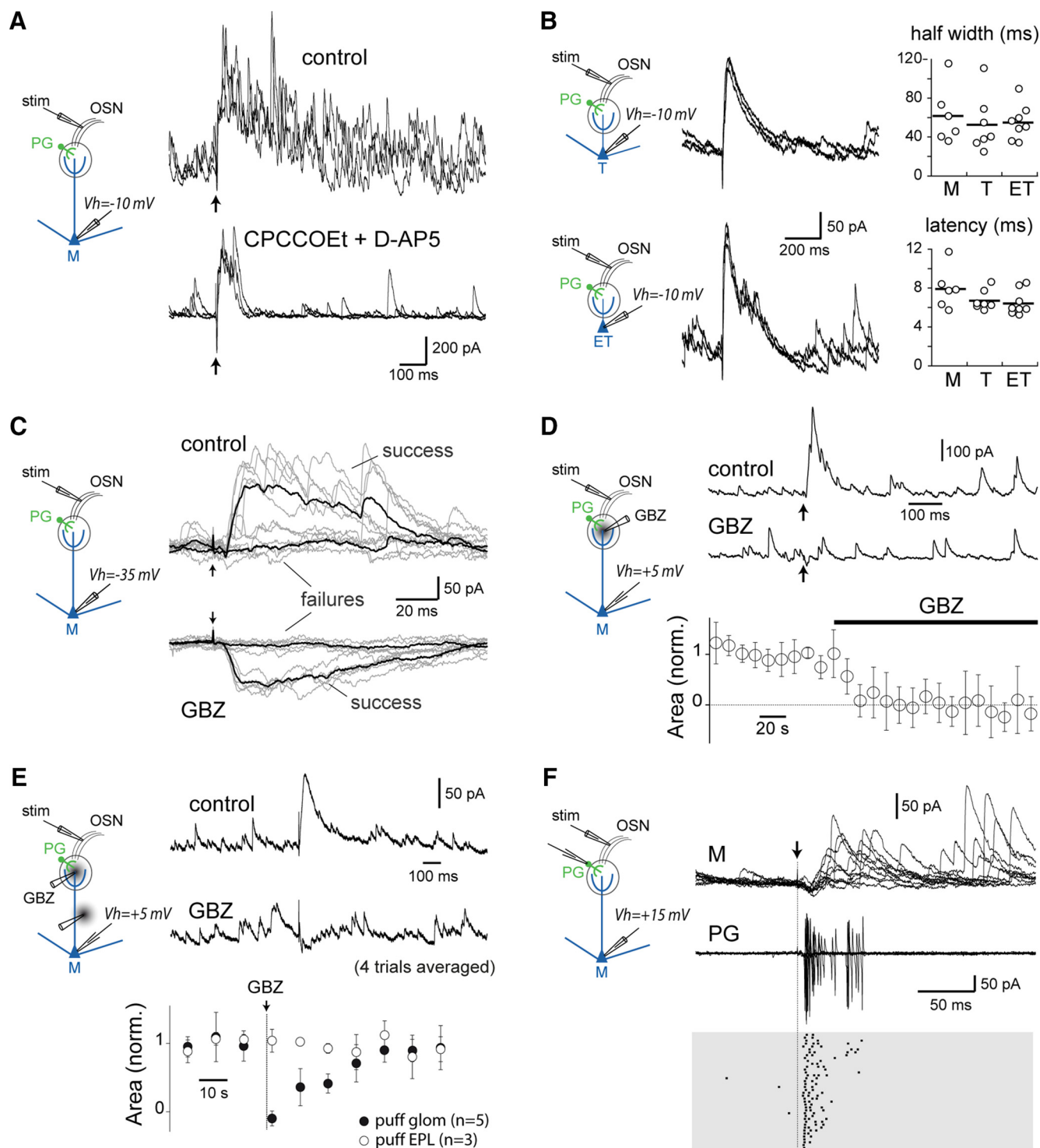
**Figure 6.** Activity of EYFP<sup>+</sup> PG cells in anesthetized mice. **A**, Picture of an EYFP<sup>+</sup> PG cell first recorded in the cell-attached mode (top) and then filled with Alexa Fluor 594 (red) during whole-cell recording (bottom). The spontaneous firing from this cell (top trace) was recorded in the cell-attached configuration before breaking the membrane. The corresponding respiration cycle is shown below the trace (upward deflection corresponds to inhalation). Respiration cycles were aligned, and PSTHs per respiration cycle were constructed (right). Bin width, 2 ms. The orange trace is the best Gaussian fit of the histogram. The average respiration cycle waveform is shown below the PSTH. Bottom, Spontaneous EPSCs recorded in the whole-cell voltage-clamp configuration. Right, Thirty consecutive respiration cycles are superimposed, as are their corresponding whole-cell recordings. The white trace is a typical event. **B**, Competing EPSCs and IPSCs underlie the spontaneous rhythmic activity of tufted cells *in vivo*. Left, Two-photon image of an EYFP<sup>+</sup> tufted cell during whole-cell recording *in vivo*. The patch pipette containing Alexa Fluor 594 (red) comes from the right. Middle traces, Slow outward inhibitory currents locked to the respiration rhythm (resp) dominated the spontaneous activity of this cell at  $V_h = 0$  mV (top), whereas slow inward excitatory currents dominated the activity at  $V_h = -80$  mV (bottom). Right, Fifty consecutive respiratory cycles with their correlated currents are superimposed. Black traces are the average.

suggest that EYFP<sup>+</sup> PG cells faithfully follow the activity of mitral and tufted cells that also fired bursts of action potentials at each respiration cycle (data not shown) as shown previously (Luo and Katz, 2001; Margrie and Schaefer, 2003). Therefore, intraglomerular lateral inhibition should be activated at each sniff and compete with the depolarization that generates the firing of principal neurons (Luo and Katz, 2001; Margrie and Schaefer, 2003). We tested this hypothesis in voltage-clamped tufted rather than mitral cells that are localized deeper in the brain and more difficult to voltage clamp. As expected, rhythmic slow IPSCs (half-width,  $88 \pm 22$  ms;  $n = 7$ ) correlated with the respiration were recorded at the reversal potential for excitation ( $V_h = 10/0$  mV). As illustrated in Figure 6B, in cells in which EPSCs and IPSCs were recorded successively, IPSCs occurred at the same frequency and at the same phase of the respiration cycle as EPSCs ( $n = 5$ ). Therefore, these data indicate that circuits mediating intraglomerular lateral inhibition operate *in vivo* in the absence of exogenous odorants at each respiration and thus likely provide a

rhythmic inhibition that competes with the excitation of mitral and tufted cells.

#### ON-evoked inhibition is generated within the glomerulus and involves EYFP<sup>+</sup> PG cells

In slices, stimulation of OSNs evokes a prominent barrage of summing discrete IPSCs in mitral and tufted cells (Najac et al., 2011). A recent study suggests that intraglomerular circuits mediate the early part of this OSN-evoked inhibition (Shao et al., 2012). Consistent with this study, the early component of the composite ON-evoked IPSC persisted in the presence D-AP-5 and CPCCOEt (onset latency,  $7.9 \pm 2.1$  ms; half-width,  $61.6 \pm 29.6$  ms;  $n = 6$  mitral cells; Fig. 7A) and was fully blocked when GBZ was applied locally, either continuously (Fig. 7D) or transiently (Fig. 7E), inside the glomerulus. In both cases, spontaneous IPSCs were not blocked. ON stimulation evoked a similar inhibition in tufted cells (onset latency,  $6.7 \pm 1.0$  ms; half-width,  $52.7 \pm 29.3$  ms;  $n = 7$ ) and ET cells (onset latency,  $6.4 \pm 1.3$  ms;



**Figure 7.** EYFP<sup>+</sup> PG cells mediate OSN-evoked feedforward inhibition. **A**, Mitral cell (M) responses evoked by a stimulation of OSNs in control conditions (top) and in the presence of CPCCOEt (100  $\mu$ M) and D-AP-5 (50  $\mu$ M; bottom). Individual traces recorded at  $V_h = -10$  mV are superimposed. **B**, ON-evoked feedforward inhibition in a tufted cell (T, top) and in an ET cell (bottom). Individual traces recorded at  $V_h = -10$  mV in the presence of CPCCOEt (100  $\mu$ M) and D-AP-5 (50  $\mu$ M) are superimposed. Right, Summary plots comparing the half-width and latency of the IPSC in mitral, tufted, and ET cells. Horizontal bars are the average. **C**, Mitral cell responses evoked by minimal stimulation of OSNs that evoked a response (success) in  $\sim 50\%$  of the trials in the presence of CPCCOEt plus D-AP-5 before (top, control) and after (bottom, 4  $\mu$ M) bath application of GBZ. Individual traces recorded at  $V_h = -35$  mV are superimposed in gray. Average responses are shown in black. **D**, ON-evoked mitral cell responses in the presence of CPCCOEt plus D-AP-5 (top) and during local perfusion of GBZ (25  $\mu$ M) inside the glomerulus to which the recorded mitral cell projected (middle trace). Single traces recorded at  $V_h = 5$  mV. Note that intraglomerular perfusion of GBZ did not block spontaneous IPSCs. Bottom, Normalized area of the evoked IPSCs (measured during the first 100 ms) over time during local perfusion of GBZ within the glomerulus. Average from seven cells (5 mitral and 2 tufted cells). **E**, Same as in **D**, but GBZ was applied transiently (50  $\mu$ M, 150 ms). Bottom, Summary plot for one cell for which GBZ was applied within the glomerulus (filled circles) or in the EPL below the glomerulus (open circles). **F**, Paired recording of a mitral cell and an EYFP<sup>+</sup> PG cell projecting into the same glomerulus. The mitral cell was recorded in the whole-cell mode ( $V_h = 15$  mV), whereas the PG cell was recorded in loose cell-attached mode. A minimal stimulation (18  $\mu$ A) was used to stimulate OSNs in the presence of CPCCOEt plus D-AP-5. IPSCs in the mitral cell (top) were correlated with action potentials in the PG cell (middle). Bottom, Raster plot of every successful trial in the PG cell.

half-width,  $54.9 \pm 17.8$  ms;  $n = 8$ ; no statistical difference with values in mitral cells; Fig. 7B). Together, D-AP-5 and CPCCOEt shorten the excitation of mitral and tufted cells (De Saint Jan and Westbrook, 2007) and, consequently, the excitatory drive of interneurons. In addition, the NMDA receptor antagonist limits GABA release from granule cells (Isaacson and Strowbridge, 1998; Schoppa et al., 1998). Because IPSCs were evoked even when OSNs were stimulated with minimal intensities ( $25\text{--}50\ \mu\text{A}$ ,  $n = 10$  mitral cells; Fig. 7C), interneurons that mediate this inhibition should be activated by weak ON inputs. Do EYFP<sup>+</sup> PG cells participate in ON-evoked inhibition? To investigate this possibility, we recorded IPSCs in a mitral cell while monitoring the activity of an EYFP<sup>+</sup> PG cell in the loose cell-attached mode under conditions of minimal stimulation that recruit only a fraction of PG cells but activate inhibition. Under these conditions and in the presence of D-AP-5 and CPCCOEt, EYFP<sup>+</sup> PG cells fired  $2.4 \pm 0.9$  action potentials ( $n = 4$  pairs) in  $95 \pm 5\%$  of the trials in which a compound IPSC was evoked in the mitral cell (Fig. 7F). Together, these results suggest that glomerular circuits mediate the early phase of ON-evoked inhibition and that EYFP<sup>+</sup> PG cells contribute to this inhibition.

### Glomerular inhibition promotes spike timing variability in mitral cells

A single stimulation of OSNs evokes polysynaptic EPSCs and IPSCs in mitral and tufted cells (Najac et al., 2011). Whereas the excitation is almost identical from trial to trial, the inhibition is composed of asynchronous discrete IPSCs with variable timing. To assess the influence of intraglomerular inhibition on the spike output of principal cells, we made loose cell-attached recordings in mitral cells and compared their ON-evoked discharge under control conditions and when intraglomerular inhibition was blocked by a local application of GBZ within the glomerulus ( $n = 9$ ; Fig. 8). In some experiments ( $n = 3$ ), we simultaneously monitored evoked IPSCs in a second cell projecting into the same glomerulus to better control the stimulation and the effect of GBZ, as in the example shown in Figure 8, B and C. Mitral cells responded to the stimulation with a long-lasting barrage of spikes that returned to baseline frequency after a double-exponential decay ( $25.3 \pm 31.4$  and  $369 \pm 184$  ms). Intraglomerular application of GBZ increased firing rate ( $32 \pm 26\%$  increase compared with control during the 3 s after the stimulus,  $p = 0.006$ ), especially during the first 200 ms after the stimulus (Fig. 8B,C). Furthermore, spike rasters of individual neurons suggest that spike time tended to be more reliable in the presence of GBZ (Fig. 8C). To quantify the effect of intraglomerular inhibition on spike timing, we ranked the interspike intervals (ISIs) in each trial for each mitral cell. We compared the distribution of their duration across trials in the presence and absence of GBZ in a short time window (200 ms) after ON stimulation (Fig. 8D). Most ISIs after the stimulus were significantly different in the two conditions for nearly all neurons (two-sampled Kolmogorov–Smirnov test,  $p \leq 6.15 \times 10^{-4}$ ; Fig. 8E). However, these differences in the ISI duration distributions could result from both increased firing rates and increased spike reliability in the presence of GBZ. Therefore, we estimated the coefficient of variation ( $CV_{\text{ISI}}$ ) of each ISI across trials in the two conditions (Fig. 8F). We found that the CV of the first two ISIs after the stimulus onset were significantly smaller in the presence of GBZ (first ISI:  $CV_{\text{ISI}}^{\text{GBZ}} = 0.11 \pm 0.06$ ,  $CV_{\text{ISI}}^{\text{CNT}} = 0.52 \pm 0.35$ ,  $p \leq 0.01$ ; second ISI:  $CV_{\text{ISI}}^{\text{GBZ}} = 0.18 \pm 0.14$ ,  $CV_{\text{ISI}}^{\text{CNT}} = 0.44 \pm 0.26$ ,  $p \leq 0.01$ ). Subsequent ISIs did not show any significant differences in CV across trials. Thus, intraglomerular inhibition not only reduces spike frequency but also

promotes trial-to-trial spike timing variability within a short time window after ON stimulation. To understand how intraglomerular lateral inhibition could affect the response of an ensemble of cells that receive a simultaneous ON input, we then simulated an ensemble of nine mitral cells by pooling the data from our individual recordings. For every ensemble response, we calculated the CV of ISIs across the nine mitral cells in the presence and absence of GBZ (Fig. 8G). We found that the CV of the first two ISIs after the stimulus onset were significantly larger in the control condition than in the presence of GBZ (first ISI:  $CV_{\text{ISI}}^{\text{GBZ}} = 0.36 \pm 0.08$ ,  $CV_{\text{ISI}}^{\text{CNT}} = 1.04 \pm 0.20$ ,  $p \leq 0.001$ ; second ISI:  $CV_{\text{ISI}}^{\text{GBZ}} = 0.24 \pm 0.08$ ,  $CV_{\text{ISI}}^{\text{CNT}} = 0.61 \pm 0.21$ ,  $p \leq 0.001$ ). This increase in cell-to-cell spike redundancy in GBZ was restricted to the first few spikes after ON stimulation as for the trial-to-trial reliability described above for each cell.

Thus, intraglomerular inhibition counteracts membrane properties and dendrodendritic electrical and synaptic interactions that constrain mitral cells to fire reliable and precisely timed action potentials in response to a stereotyped excitatory input, thereby reducing the similarity of the discharge of each cell.

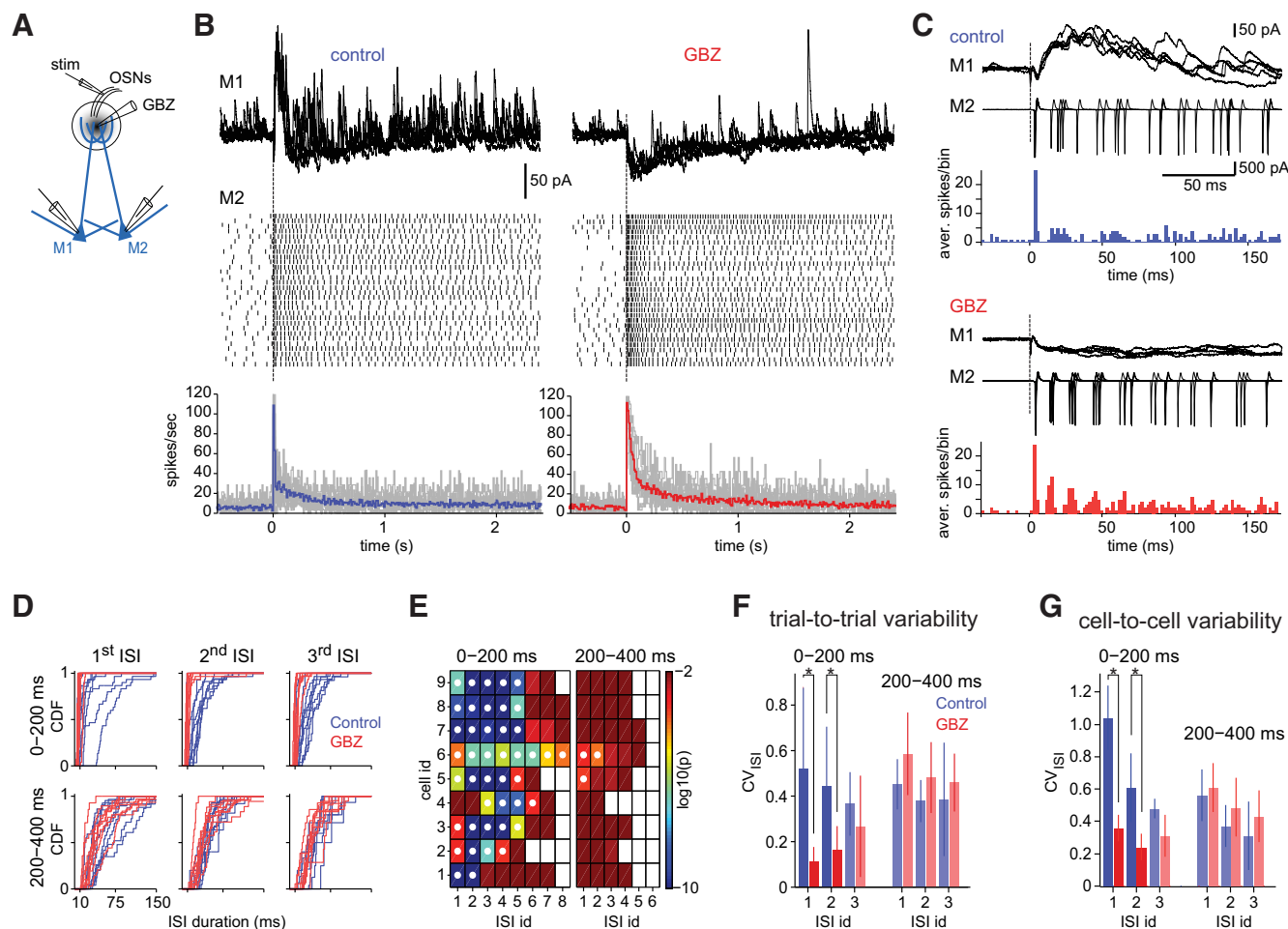
### Discussion

We characterized a specific population of PG cells that mediates lateral inhibition between olfactory bulb principal neurons projecting into the same glomerulus. This intraglomerular inhibitory pathway is recruited by even the weakest ON input and shapes the spike output of mitral and tufted cells. Moreover, *in vivo* recordings show that intraglomerular lateral inhibition is activated at each respiration cycle.

We took advantage of a transgenic mouse expressing EYFP in a subset of PG cells to characterize the properties of an intraglomerular inhibitory circuit that modulates the spike output of principal neurons. EYFP<sup>+</sup> PG cells include, although not exclusively, cells expressing CB but does not overlap with the two other well identified populations of PG cells immunoreactive for CR and TH. Electron microscopy studies of CB<sup>+</sup> PG cells have shown that they receive synapses from mitral/tufted cells but rarely from OSNs (Toida et al., 1998; Kosaka and Kosaka, 2007; Panzanelli et al., 2007). Moreover, ultrastructural evidence suggests that CB<sup>+</sup> PG cells do not have an axon and release their neurotransmitter from dendrites within their glomerulus (Kosaka and Kosaka, 2010). Our functional and morphological characterization of EYFP<sup>+</sup> PG cells is consistent with these data. We have demonstrated that mitral, tufted, and ET cells make excitatory synapses on EYFP<sup>+</sup> PG cells. Thus, EYFP<sup>+</sup> PG cells responded to a stimulation of OSNs with a short barrage of EPSCs that lasted only few tens of milliseconds. Similar responses were observed in a subgroup of EYFP<sup>−</sup> PG cells, suggesting that EYFP<sup>+</sup> PG cells, although they constitute 30% of PG cells, might be representative of an even larger group of type 2 PG cells with similar excitatory connections. The transient nature of this response, which contrasts with the long-lasting responses observed in other PG cells, was remarkably consistent despite the diversity of immunohistochemical profiles and membrane properties among EYFP<sup>+</sup> PG cells. The onset latency of the response was quite fast ( $\sim 3$  ms) yet longer and more variable than in ET or PG cells connected directly to OSNs, suggesting that EYFP<sup>+</sup> PG cells do not receive direct inputs from OSNs. This fast-onset polysynaptic activation likely relies on ET cells that fire as early as 2 ms after the stimulation of OSNs (De Saint Jan et al., 2009) and elicit an EPSC in EYFP<sup>+</sup> PG cells in  $\sim 1$  ms.

Some PG cells are activated by weak ON inputs (Gire and Schoppa, 2009) or the firing of a single principal cell (Murphy et





**Figure 8.** Intraglomerular inhibition reduces output frequency and spike timing redundancy in mitral cells. **A**, Schematic of the experiment. **B**, Paired recording of ON-evoked responses in two mitral cells (M1 and M2) projecting into the same glomerulus. M1 (top traces) was recorded in whole-cell mode at  $V_h = 5$  mV. Five responses are superimposed under control conditions (left) and during local perfusion of GBZ ( $25 \mu\text{M}$ ) within the glomerulus (right). Middle, Raster plots of the firing of M2 recorded simultaneously in loose cell-attached mode. The dashed line indicates the stimulation ( $25 \mu\text{A}$ ) time. Bottom, Mean firing rate across nine mitral cells recorded in cell-attached mode in control conditions (left, blue) and in the presence of GBZ (right, red). Thick traces are grand averages, and gray traces are averages for each cell. Bin width, 10 ms. **C**, Zoom on the initial phase of the responses shown in **B**. Bottom, PSTH for the same M2 cell in control conditions (top) and in the presence of GBZ (bottom). Bin width, 2 ms. **D**, Top, Cumulative density function (CDF) of the first three ISIs (first 4 spikes) in the 0–200 ms time window for nine mitral cells recorded in cell-attached mode. Blue colors correspond to the control data, and red colors indicate GBZ application. The three ISIs are shown in different columns. Bottom, Same as the top for the 200–400 ms time window. **E**, Significance testing of the difference between ISI distribution in control conditions and in the presence of GBZ. Two-sampled Kolmogorov–Smirnov test  $p$  values are shown in log scale. Null hypothesis is that the ISI distribution in control conditions and in the presence of GBZ are the same.  $x$ -Axis refers to the ISI, and the  $y$ -axis refers to the cell number.  $p$  values are higher within the 0–200 ms than in the 200–400 ms time period. Significantly different ISIs in control and GBZ are marked with a white dot ( $\alpha$  values = 0.001,  $n = 9$  mitral cells). **F**, In the presence of GBZ, the first two ISIs after stimulation become highly reliable across trials.  $\text{CV}_{\text{ISI}}$  of the first three ISIs. In **F** and **G**, each bar shows the mean  $\pm$  SD of  $\text{CV}_{\text{ISI}}$  in control conditions (blue) and in the presence of GBZ (red) for the first three ISIs within the 0–200 and 200–400 ms time windows. **G**, In the presence of GBZ, in each trial, the first two ISIs after stimulation become highly reliable across cells in a simulated ensemble of nine mitral cells ( $n = 14$  random trials).

al., 2005). Current models suggest that low-threshold intraglomerular inhibition may serve as a gatekeeper that shunts weak ON inputs (Gire and Schoppa, 2009) and may separate the firing phase of the two output channels (tufted vs mitral cells) along a respiration cycle (Fukunaga et al., 2012). Our results show that  $\text{EYFP}^+$  PG cells constitute a large fraction of these highly excitable PG cells that also include a minority of  $\text{EYFP}^-$  PG cells. We have demonstrated that they mediate a form of lateral inhibition, the firing of a single principal neuron producing disinhibitory inhibition of other mitral and tufted cells within the same glomerulus. This recurrent pathway is recruited by even the weakest ON input and mediates ON-evoked inhibition of principal neurons. We also found that  $\text{EYFP}^+$  PG cells indiscriminately release GABA onto mitral, tufted, and ET cells and that ON-evoked intraglomerular inhibition has a similar time course in these three types of neurons. Our experimental results do not support a re-

cent study that stated that, unlike mitral cells, tufted cells are not, or are less, modulated by intraglomerular inhibition (Fukunaga et al., 2012). However, that study relied on a theoretical network model based on experimentally undemonstrated differences in PG cell inputs on mitral and tufted cells. More recently, Burton and Urban (2014) suggested that the different firing phases of mitral and tufted cells may be attributable to a greater intrinsic excitability and a stronger afferent-evoked excitation in tufted cells.

Evaluating the contribution of  $\text{EYFP}^+$  PG cells in ON-evoked inhibition would require their selective manipulation, for instance, with optogenetics. However, to date, there is no selective marker for this population, which impedes its genetic manipulation. Nevertheless, our data suggest that only a minority of PG cells have a similar activation threshold than  $\text{EYFP}^+$  PG cells and potentially contribute to ON-evoked inhibition. Moreover, we

show that half of CR<sup>+</sup> PG cells are not, or are only weakly, activated by ON inputs and thus unlikely to participate. However, ON-evoked inhibition lasts longer than the firing of EYFP<sup>+</sup> PG cells, suggesting that other PG cell subtypes that respond to an ON input with a longer firing may generate late IPSCs. Do granule or PV<sup>+</sup> cells also contribute to the early phase of ON-evoked inhibition? Several results argue against this idea. As shown previously (Shao et al., 2012), early IPSCs persisted in the presence of NMDA receptor blockers that limit the activation of granule cells (Isaacson and Strowbridge, 1998; Schoppa et al., 1998). Second, ON-evoked inhibition was abolished by intraglomerular perfusion of GBZ, suggesting that PV<sup>+</sup> and granule cells that do not make synapses within the glomerulus do not contribute. Consistent with this conclusion, the time course of ON-evoked inhibition was similar in ET cells that presumably do not interact with granule cells and PV<sup>+</sup> interneurons. This indicates that intraglomerular pathways are sufficient to produce a strong inhibition. Finally, granule cells respond to a glomerular stimulation with an ~70 ms delayed firing (Schoppa and Westbrook, 1999; Kapoor and Urban, 2006) that does not match with the early onset of OSN-evoked inhibition (~6 ms).

The short timescale of EYFP<sup>+</sup> PG cell-mediated intraglomerular inhibition in slices (~100 ms) is physiologically relevant for odor processing. *In vivo*, spontaneous firing and odor-evoked discharges of mitral and tufted cells are often constrained by the ~2–10 Hz respiratory rhythm. Thus, mitral and tufted cells fire action potentials during a brief (50–150 ms) phase of the respiration cycle (Cang and Isaacson, 2003; Margrie and Schaefer, 2003; Davison and Katz, 2007; Dhawale et al., 2010). As reported previously (Luo and Katz, 2001; Margrie and Schaefer, 2003), rhythmic depolarizations locked to the respiration underlie this activity. We have found that these EPSPs compete with IPSPs in anesthetized mice. Because EYFP<sup>+</sup> PG cells fire action potentials at each respiration cycle, they likely mediate at least a fraction of this inhibition. Similar to EYFP<sup>+</sup> PG cells, PV<sup>+</sup> cells are highly active *in vivo*, even in the absence of exogenously applied odorants (Miyamichi et al., 2013), and thus may contribute to this inhibition. In contrast, granule cells have a low spontaneous firing rate in anesthetized mice (Cang and Isaacson, 2003; Margrie and Schaefer, 2003; Kato et al., 2012). However, their activity is strongly enhanced in the awake animal, suggesting that the influence of each interneuron population may differ in the awake and anesthetized state (Kato et al., 2012).

In classical pathways, feedforward inhibition quickly follows monosynaptic excitation and defines a short temporal window of opportunity for the postsynaptic neuron to fire. Thus, inhibition often promotes spike timing precision (Pouille and Scanziani, 2001). In contrast, we found that ON-evoked intraglomerular inhibition promotes spike timing variability in olfactory bulb mitral cells from trial to trial and also from cell to cell. Intraglomerular lateral inhibition relies on synapses with low release probability, whereas the long-lasting ON-evoked excitation of mitral and tufted cells relies on excitatory synapses with high release probability (Najac et al., 2011). Therefore, IPSC timing and amplitudes vary from trial to trial and from cell to cell, unlike excitation that is highly reproducible. Moreover, membrane properties constrain mitral cells to fire reliable and precisely timed action potentials across trials in response to an identical synaptic-like fluctuating stimulus (Padmanabhan and Urban, 2010) or an EPSP waveform (Balu et al., 2004). This explains why each mitral cell responded to the afferent input with a stereotyped pattern of spikes in the presence of GBZ. In addition, intraglomerular dendrodendritic excitatory interactions and

gap-junction coupling drive synchronous firing in principal cells connected to the same glomerulus (Schoppa and Westbrook, 2002; Ma and Lowe, 2010). Thus, intraglomerular inhibition competes with multiple mechanisms that tend to synchronize the output of a glomerulus. Many neurons of the olfactory bulb network fire in synchrony in the presence of an odorant, suggesting that olfactory coding in the olfactory bulb relies, at least in part, on precise spike timing in a population of cells (Bathellier et al., 2010). Although the functional implication of intraglomerular inhibition is at present uncertain, one possible function of this pathway may be to prevent correlated spiking in the absence of exogenous odorants.

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